Dynamics of Enhanced Tracer Diffusion in Suspensions of Swimming Eukaryotic Microorganisms

K. C. Leptos
Jeffrey S. Guasto
Jerry P. Gollub
A. I. Pesci

Follow this and additional works at: https://scholarship.haverford.edu/physics_facpubs

Repository Citation

This Journal Article is brought to you for free and open access by the Physics at Haverford Scholarship. It has been accepted for inclusion in Faculty Publications by an authorized administrator of Haverford Scholarship. For more information, please contact nmedeiro@haverford.edu.
Dynamics of Enhanced Tracer Diffusion in Suspensions of Swimming Eukaryotic Microorganisms

Kyriacos C. Leptos,1 Jeffrey S. Guasto,2 J. P. Gollub,1,2 Adriana I. Pesci,1 and Raymond E. Goldstein1

1Department of Applied Mathematics and Theoretical Physics, University of Cambridge, Wilberforce Road, Cambridge CB3 0WA, United Kingdom
2Department of Physics, Haverford College, Haverford, Pennsylvania 19041, USA

(Received 25 June 2009; revised manuscript received 20 October 2009; published 5 November 2009)

In contexts such as suspension feeding in marine ecologies there is an interplay between Brownian motion of nonmotile particles and their advection by flows from swimming microorganisms. As a laboratory realization, we study passive tracers in suspensions of eukaryotic swimmers, the alga Chlamydomonas reinhardtii. While the cells behave ballistically over short intervals, the tracers behave diffusively, with a time-dependent but self-similar probability distribution function of displacements consisting of a Gaussian core and robust exponential tails. We emphasize the role of flagellar beating in creating oscillatory flows that exceed Brownian motion far from each swimmer.

DOI: 10.1103/PhysRevLett.103.198103

PACS numbers: 47.63.Gd, 05.40.—a, 87.17.Jj, 87.23.—n

For many of the microscopic organisms in marine and lake environments the process of feeding involves an interplay between diffusion of target particles and their advection by flows from swimming microorganisms. As a laboratory realization, we study passive tracers in suspensions of eukaryotic swimmers, the alga Chlamydomonas reinhardtii. While the cells behave ballistically over short intervals, the tracers behave diffusively, with a time-dependent but self-similar probability distribution function of displacements consisting of a Gaussian core and robust exponential tails. We emphasize the role of flagellar beating in creating oscillatory flows that exceed Brownian motion far from each swimmer.

Here we report an advance in the study of tracer statistics in suspensions of swimmers. First, we use the alga Chlamydomonas reinhardtii [11], a biflagellated model eukaryote in the study of photosynthesis and flagella. Relative to the bacteria studied previously [6] (E. coli, B. subtilis) Chlamydomonas has features that will greatly simplify the theoretical interpretation of our results; it is nearly spherical, is sufficiently large (a radius $R \sim 5 \mu$m) that it is not subject to substantial rotational diffusion, and as its two 10–12 μm long flagella beat in a synchronized “breast stroke” pattern at $\sim$50 Hz, it displays long periods of nearly straight-line motion (up to $\sim 10$ s [12]). Second, we characterize in detail the time-dependent PDF of tracer displacements, and find clear systematic deviations from Gaussianity, in the form of an exponential tail whose amplitude grows with swimmer concentration. Despite this complex form, the PDFs exhibit self-similarity under a diffusive scaling. Some physical considerations that may be important in a theory of these results are suggested.

Microscopy of Chlamydomonas suspensions was carried out on a Nikon TE2000-U inverted fluorescence microscope inside disk-shaped chambers fabricated from polydimethylsiloxane (PDMS), 1 cm in diameter and 1.5 mm deep. After casting, each cavity was etched in an oxygen plasma (Femto, Diener Electronic) and bonded to a PDMS-spin-coated cover slip. Both the interior of the chambers and the fluorescent particles were pacified using a 5% (w/v) solution of bovine serum albumin to prevent the protein in the flagella from adhering to them [13]. Chlamydomonas reinhardtii (UTEX 89) was grown axenically in Tris-Acetate-Phosphate (TAP) medium [11], on an orbital shaker (300 rpm) in a diurnal growth chamber (KBW400, Binder). The daily cycle was 16 h of cool white
light (∼80 μmol photons/m² PAR) at 28 °C, and 8 h in the dark at 26 °C. For concentrations >10⁶ cells/cm³, cells were concentrated from exponentially growing cultures by centrifugation in tubes at 1100 g and addition of fresh TAP to the cell pellet. The tubes were then placed in the diurnal chamber for at least 1 h to allow the cells to recuperate. All cell concentrations were measured using a haemocytometer [14]. Fluorescent microspheres (yellow-green, Invitrogen F8827; radius a = 1 μm) were added to obtain a final concentration of ∼250 ppm (w/v). Each sample was transferred to a separate PDMS observation chamber, taking care to avoid excessive shear that might damage the flagella. Movies of cells and of tracer particles were recorded with a high-speed video camera (Fastcam SA-3, Photron) at up to 500 fps to resolve motion induced by flagellar beating. Tracers were observed at ×40 (NA 0.6) with epifluorescence illumination (Pantachrome, Till Photonics) in a plane ∼100 μm above the chamber bottom. Cell and particle paths were extracted from recordings with a predictive particle tracking algorithm [15].

A relatively dilute suspension of cells (5 × 10⁶ cm⁻³) in growth medium was first studied at ×20 magnification (NA 0.45) with bright-field illumination, using a red long-pass filter (λ > 620 nm) to prevent phototaxis. While cell trajectories have been studied previously [12], it is important to characterize them in situ along with the tracer dynamics. The representative sample in Fig. 1(a) illustrates a broad spectrum of trajectories including helical paths of variable pitch—the well-known spinning of the body around the trajectory midline [11]. The cell speed distribution in the imaging plane [Fig. 1(b)] shows a peak at ∼100 μm/s. There is some out-of-plane motion that is not imaged, so the 3D distribution may be even sharper. The distribution is consistent with swimming at approximately constant speed in random directions. Indeed, while many trajectories’ centerlines are gently curved, sharp turns are only observed for Δt > 10 s [12].

A statistical analysis of cell displacements along an arbitrary axis in the imaging plane shows ballistic behavior [Fig. 1(b), inset] up to several seconds. Tracer trajectories [Fig. 1(c) and inset] involve both Brownian components and large jumps induced by flows from the swimmers. Higher magnification (inset) shows that these advective motions are loops due to flagellar beating.

For each volume fraction ϕ of Chlamydomonas the PDF of tracer displacements P(Δx, Δt) along an arbitrary direction was calculated for a number of increasing time intervals Δt. The intervals used were well within the ballistic regime of the swimmers [Fig. 1(b)], and an order of magnitude smaller than the mean time between sharp turns measured in three-dimensional tracking experiments [12]. Figure 2(a) shows the PDF at a fixed interval (Δt = 0.12 s) for increasing ϕ. Without swimmers, the distribution is accurately fitted by a Gaussian whose width is determined by the diffusion constant D₀ = 0.28 μm²/s, consistent with the Stokes-Einstein value.

In the presence of swimmers, there are two changes to the distributions. First, exponential tails appear in the PDFs, the magnitudes of which grow with increasing concentration of swimmers. Second, the Gaussian core broadens significantly. For all volume fractions ϕ and time intervals Δt the distributions P(Δx, Δt) can be fitted approximately (in log(P), using a nonlinear least squares algorithm) to a weighted sum of Gaussian and exponential (Laplace) distributions,

\[
P(\Delta x, \Delta t) = \frac{1 - f}{(2\pi \delta_x^2)^{1/2}} e^{-(\Delta x^2)/2\delta_x^2} + \frac{f}{2\delta_x} e^{-|\Delta x|/\delta_x}. \tag{1}
\]

The mean absolute deviation between the fit and the data for log(P) is ∼0.1, corresponding to 3% of the range of log(P) shown in the data. There are three parameters: the standard deviation δ_x(Δt, ϕ) of the Gaussian distribution, the decay length δ_x(Δt, ϕ) of the Laplace distribution, and the fractional contribution f(ϕ) of advection-enhanced displacements. We find that both lengths exhibit diffusive growth, with δ_x ≈ A_x(Δt)¹/₂ and δ_x ≈ A_x(Δt)¹/₂. This im-

**FIG. 1** (color online). Swimming cells and tracer particles. (a) *Chlamydomonas* trajectories over an interval of ∼1 s. (b) PDF of swimming speed obtained at 30 fps, averaged over 30 s, and (inset) mean squared displacement of swimmers in the imaging plane. The line (Δx²) ~ (Δt)² shows the motion to be ballistic for intervals up to several s. (c) Trajectories of 2.0 μm microspheres observed at 50 fps. Inset: loopy tracer path, sampled at 500 fps (bright-field), induced by a cell passing within ∼8 μm.
grounds we expect

(b) Diffusive rescaling of PDFs at 

time intervals, illustrating data collapse.

plies that a diffusive rescaling of the displacements in terms of $\Delta x/(2D_{\text{eff}}\Delta t)^{1/2}$ leads, at any given $\phi$, to a data collapse, as shown in Fig. 2(b), where $D_{\text{eff}}$ is a constant (see below). Furthermore, despite the complex shape of the PDFs, the mean square displacement of the tracers remains linear in time, with a slope that increases with $\phi$ [Fig. 3(a)], apart from small variations from run to run that are not statistically significant. Similar trends have been seen in thin films of swimming bacteria [6]. We characterize this behavior by means of an effective diffusivity $D_{\text{eff}} = \langle(\Delta x)^2 \rangle/2\Delta t$, and find that $D_{\text{eff}}$ increases with the volume fraction of swimmers. These measurements suggest a heuristic picture of the suspension; surrounding each cell is a sphere of influence of radius $R_{\text{eff}}$ within which advection strongly dominates diffusion, and outside of which tracers experience only effective Brownian motion. This is consistent with the PDFs having the form of a weighted sum of the two distributions. $R_{\text{eff}}$ and $f$ provide measures of this sphere size; at the highest concentration ($\phi = 2.2\%$, $f \approx 0.12$) the mean distance between spheres is $\sim 30 \mu$m, almost thrice $R_{\text{eff}}$, so binary interactions are negligible.

A quantitative measure of the advection/diffusion balance comes from considering the flow field around a cell in the simplified form $u(r) \exp(i\omega t)$. Tracer oscillations are indistinguishable from Brownian motion at a radius $r_B > R_{\text{eff}}$ where the displacement $2u(r_B)/\omega$ in a half cycle is comparable to the diffusive displacement $(2D t)^{1/2}$ in a time $t = \pi/\omega$. For *Chlamydomonas* flagellar beating with $\omega = 2\pi \times 50 \text{ rad/s}$ this yields $u(r_B) \sim (2\pi D \omega)^{1/2} \sim 10 \mu$m/s, and a displacement $\sim 0.1 \mu$m. A direct experimental determination of $r_B$ can be obtained from simultaneous bright-field microscopy of tracer motion and free-

![FIG. 2 (color online). Probability distribution functions for tracer displacements. (a) Overlaid PDFs at a fixed time interval $\Delta t = 0.12$ s, for various volume fractions, showing the broadening of the Gaussian core and appearance of exponential tails. (b) Diffusive rescaling of PDFs at $\phi = 2.2\%$ and various time intervals, illustrating data collapse.](image)

![FIG. 3 (color online). Diffusional properties. (a) Mean squared tracer displacement at various cell concentrations. Concentration dependence of (b) effective diffusivity, with linear fit, and (c) fractional contribution of PDF tails.](image)
swimming cells. Figure 4(a) shows one such trajectory, under the same conditions as the inset of Fig. 1(c), plotted in the frame of reference of the swimming *Chlamydomonas*, along with the time series of radial velocity in the laboratory frame [Fig. 4(b)]. Inspection of Fig. 4(b) suggests the estimate $r_B \approx 25-30 \mu m$. For comparison one might ask when oscillatory displacements are comparable to the tracer radius, $2u(r_a)/\omega = a$, which yields $u(r_a) \sim 150 \mu m/s$. Such velocities appear at distances of order $15 \mu m$ ($\sim 1.5R_{eff}$). To go beyond these measurements to a model for the velocity field we note that the viscous penetration depth associated with the flagellar beating is $\xi = (2\nu/\omega)^{1/2} \approx 80 \mu m$. This is only a few times larger than $r_B$ so these flows are intrinsically unsteady. Detailed comparison [17] with tracer trajectories shows quantitative consistency with such models and confirms the length scale $r_B$. Moreover, the number of flagellar beats during the intervals $\Delta t$ of the PDFs, and also during the encounter time of a tracer with a swimmer, can easily be ten or more. Such unsteadiness may alter significantly encounter rates in suspension feeding relative to those for steady flow [2,3]. It also contributes to an enhanced tracer diffusivity, as in the problem of pollutant dispersal in oscillatory tidal flows [18].

We have shown that the statistics of passive tracers in suspensions of flagellated eukaryotes exhibits a non-Gaussian yet self-similar form, with clear systematics in the concentration of swimmers. Oscillatory components appear to play a significant role in the dynamics. Development of a statistical theory of the PDFs and the diffusional enhancement is clearly the outstanding theoretical challenge. In closing, we note that in contrast to a bacterium, whose helical flagella push it through the fluid, *Chlamydomonas* is a “puller”. Theoretical studies of instabilities in concentrated suspensions of swimmers [7] suggest a profound difference between the two, with only pushers displaying large-scale coherence. The techniques outlined here may allow for sufficiently high concentrations of cells to test this prediction in detail.

We thank K. Drescher, B. Eckhardt, T. J. Pedley, and C. A. Solari for discussions, and N. Damean, D. Page-Croft, and N. Price for technical assistance. Work supported by the BBSRC, the Leverhulme Trust (J.P.G.), NSF Grant No. DMR0803153 (J. S. G. & J. P. G.), and the Schlumberger Chair Fund. J. P. G. appreciates the hospitality of DAMTP during this work.

![Figure 4](image-url)

**FIG. 4** (color online). Tracer dynamics. (a) Trajectory of a tracer particle imaged at 500 fps, in a frame of reference moving with a *Chlamydomonas* cell. Black disk represents the approximate size of the cell. (b) Induced radial motion fades into Brownian motion beyond $\sim 25 \mu m$ from the cell.